Cancer risk from exposure to galactic cosmic rays: implications for space exploration by human beings

Francis A Cucinotta, Marco Durante

Space programmes are shifting toward planetary exploration, and in particular towards missions by human beings to the moon and Mars. However, exposure to space radiation is an important barrier to exploration of the solar system by human beings because of the biological effects of high-energy heavy ions. These ions have a high charge and energy, are the main contributors to radiation risk in deep space, and their biological effects are understood poorly. Predictions of the nature and magnitude of risks posed by exposure to radiation in space are subject to many uncertainties. In recent years, worldwide efforts have focussed on an increased understanding of the oncogenic potential of galactic cosmic rays. A review of the new results in this specialty will be presented here.

National Aeronautics and Space Administration, Lyndon B Johnson Space Centre, Houston, TX, 77058, USA (F A Cucinotta PhD); **and Department of Physics and National Institute of Nuclear Physics, University Federico II, Naples, Italy** (Prof M Durante PhD)

Correspondence to:
Dr F A Cucinotta
Francis.A.Cucinotta@nasa.gov

Introduction

Space exploration is an adventure for humankind, with the potential for discoveries that capture our imaginations and benefit society. The benefits from exploration[1] must be balanced with the cost, safety, and ethical concerns when deciding acceptable risks for astronauts. The main health concerns are exposure to galactic cosmic rays (GCR) and solar proton events, which lead to substantial, but poorly understood, risks of carcinogenesis and degenerative disease.[2,3] Spaceflights in low Earth orbit, such as missions on a space shuttle and at an international space station, are partly protected by the Earth's magnetic field and the solid shielding of the planet. The Apollo space missions ventured away from the protection of the Earth, but lasted only up to 12 days. Proposed missions to the moon (figure 1) in the next decade could last up to 200 days. Furthermore, a possible mission to Mars that could last up to 3 years would lead to whole-body doses of radiation of about 1 **Sievert** (Sv) or more.[4] However, the ideas used for prediction of risk on Earth, including use of the dose unit Sv, are perhaps deceptive for GCR exposure. This Essay discusses efforts to improve the understanding of biological effects of densely ionising heavy ions through biomedical research of cancer.

Space radiation environments and risk assessment

In space, astronauts are exposed to: protons; high-energy heavy (HZE) ions that have a high charge (Z) and energy (E); and secondary radiation, including neutrons and recoil nuclei produced by nuclear reactions in spacecraft walls or in tissue. The energy spectrum of GCR peaks near 1000 MeV per **nucleon**, and these particles are so penetrating that shielding can only partly reduce the doses absorbed by the crew. Thick shielding has problems for spacecraft launch systems because of its mass, and would only reduce effective GCR dose by no more than 25% with aluminium or by about 35% with the more-efficient polyethylene. Present shielding approaches cannot be regarded as a solution for the issue of radiation exposure in space, with the exception of solar proton events, which are effectively absorbed by shielding.[4]

On travelling to Mars, every cell nucleus in an astronaut's body would be hit by a proton or secondary electron (eg, electrons of the target atoms ionised by the HZE ion) every few days and by an HZE ion about once a month.[5] Whole-body doses of 1–2 mSv per day accumulate in interplanetary space and about 0·5–1 mSv per day on planetary surfaces.[6] The high ionisation power of HZE ions makes them the main contributor to risk, despite the low frequency at which they might hit a cell nucleus compared with protons. To undertake ground-based research into space radiation, special facilities are needed to accelerate charged particles (from protons to iron) to very high energies. Only a few such facilities exist in the world, and the National Aeronautics and Space Administration (NASA) has invested in a new facility at Brookhaven National Laboratory, Long Island, NY, USA.

On Earth, radiation workers or patients are most frequently exposed to low-linear-energy-transfer (LET)

Sievert
Unit of equivalent dose; Sv; m² s⁻²
1 Gy corresponds to 1 Sv for x-rays and to 20 Sv for heavy ions

Nucleon
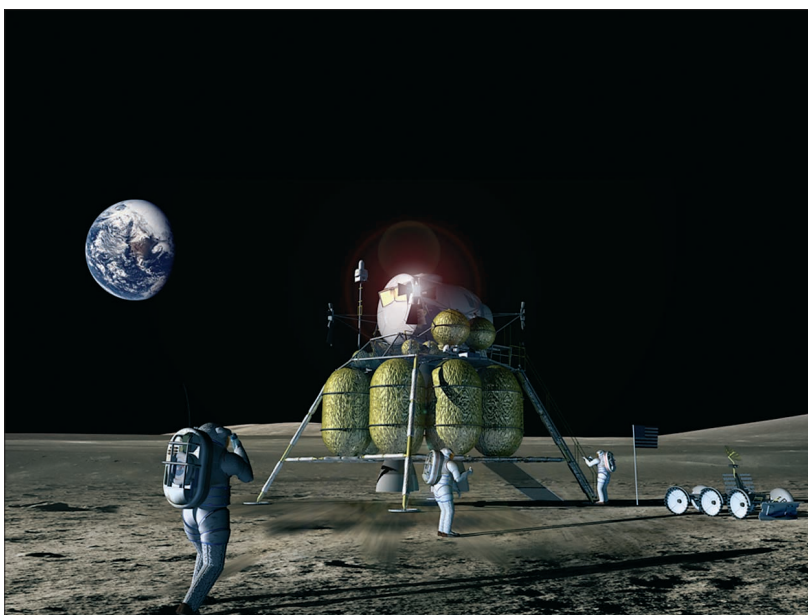A proton or neutron, the total of which make up the atomic weight

Figure 1: **A future moon landing**
According to the new Vision for Space Exploration (January, 2004), the National Aeronautics and Space Administration (NASA) plans to return to the moon in 2020. The present project anticipates four to six crew members who will complete lunar-surface exploration for 60–180 days. The Earth-moon cruise lasts about 4 days.

	Absorbed dose (Gy)*	Effective dose (Sv)	Fatal risk, % (95% CI)	
			Men (age 40 years)	Women (age 40 years)
Lunar mission (180 days)	0·06	0·17	0·68% (0·20–2·4)	0·82% (0·24–3·0)
Mars orbit (600 days)	0·37	1·03	4·0% (1·0–13·5)	4·9% (1·4–16·2)
Mars exploration (1000 days)	0·42	1·07	4·2% (1·3–13·6)	5·1% (1·6–16·4)

Calculations are at solar minimum, where GCR dose is highest behind a 5 g/cm² aluminium shield. *Mean for tissues known to be sensitive to radiation and at risk of cancer[2] including lung, colon, stomach, bladder, bone marrow, and breast and ovaries in women.[2,3] Competing causes of death are included in calculations because they decrease risk probabilities if high (ie, >5%).

Table: Radiation risks for men and women on missions to the moon or Mars

Scaling variables
Variables used to scale from atomic-bomb survivors to astronauts

Lifetable
Provides a rate of death for every year of age

radiation such as γ-rays or x-rays. Epidemiological data, mainly from survivors of the atomic bomb in Japan,[7] enable risk estimation of low-LET (ie, sparsely ionising) radiation. However, because no data for exposure to protons and HZE ions exist in human beings, risk estimates for exposure to GCR must rely entirely on experimental model systems and biophysical calculations. At present, predictions are made by use of the double-detriment **lifetable** for an average population such as that of the USA, which consists of age-dependent and sex-dependent rates of death combined with a model of radiation-induced cancer-mortality rates.[2,3] The model used for cancer mortality from radiation is based on

studies of survivors of atomic bombs,[7] which are assumed to be scalable to other populations, dose rates, and radiation types. Two **scaling variables** with large uncertainties are: the radiation-quality factor, Q, which estimates the increased effectiveness of HZE ions compared with γ-rays for the same dose; and the dose and dose-rate effectiveness factor (DDREF), which reduces estimates of cancer risk at high doses and dose-rates when dose-rates are low (ie, <0·05 Gy/h).

The table shows risks for extended missions to the moon and Mars; in this table, 95% CI are reported that account for uncertainties[3] in epidemiological data, space environments, radiation quality, and DDREF. Maximum acceptable levels of risk for astronauts are typically set at 3% fatal risk (eg, risk of mortality from cancer),[2,3] but the large uncertainties in predictions and the likelihood of other fatal or morbidity risks (eg, risk of disease) for degenerative diseases precludes whether or not a mission can be made to Mars. Use of data from survivors of atomic bombs to scale mortality for radiation risk to astronauts in space introduces many uncertainties[3] into risk estimates, and important questions remain with respect to the accuracy of any scaling approach because of qualitative differences in the biological effects of HZE ions and γ-rays.
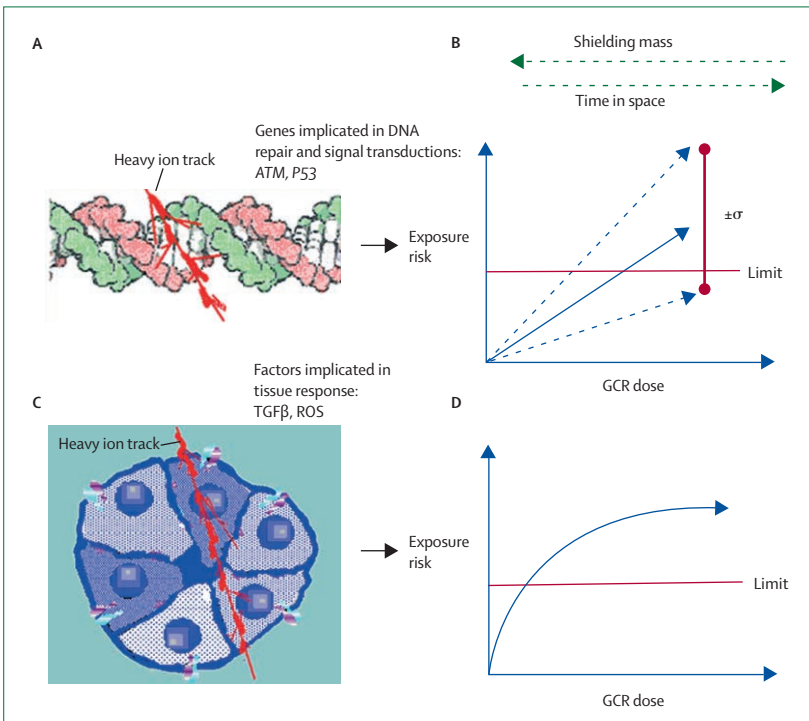
Radiobiology of HZE ions: cellular effects

A necessary step for reducing uncertainties in risk assessment are studies of molecular pathways of cancer initiation and progression, and to extend these studies to learn how such pathways can be disrupted by HZE ions including induction of genetic and epigenetic changes (figure 2). The aim of this research is to establish a more-mechanistic approach to risk estimation, and to answer questions such as: whether HZE effects can be scaled from those of γ-rays; whether risk is linear with low dose-rate; and how individual radiation sensitivity affects risk for astronauts—a population selected for many factors related to excellence in health.

First, we can analyse the initial biophysical events caused by HZE tracks (eg, a heavy ion moves along a straight track in the target material; figure 3) in cells and tissue.[6,8,9] Energy deposition by HZE ions is highly heterogeneous, with a localised contribution along the trajectory of every particle and lateral diffusion of energetic electrons (ie, δ-rays, the target atom electrons ionised by the incident HZE ion and emitted at high energy) many microns from the path of the ions. These particles are therefore densely ionising (high-LET) along the primary track (eg, the track followed by the incident heavy ion); however, they have a low-LET component because of the high-energy electrons ejected by ions as they traverse tissue. Biophysical models have shown that energy-deposition events by high-LET radiation produce different DNA lesions, including complex DNA breaks, and that qualitative differences between high-LET radiation and low-LET radiation affect both the induction

Figure 2: **Importance of uncovering basic mechanisms of cancer induction by GCR**
Defining the role of DNA damage versus that of non-targeted effects has implications for radiation shielding, mission duration, and design of biological countermeasures. In the DNA-target model (A), a linear response (B) for risk is expected, with research focus on the slope of response as a function of radiation quality and radiation sensitivity ±σ=SE on the slope of the dose-risk relation. Limit=administrative limit that should not be exceeded by astronauts. In the non-targeted model (C), shielding is ineffective in tissue (D) and distinct targets for biological countermeasures are pursued. ATM=ataxia telangiectasia mutated. TGFβ=transforming growth factor β. ROS=reactive oxygen species.

and repair of DNA damage.[10–13] The number of DNA single-strand breaks and double-strand breaks produced by radiation varies little with radiation type.[8,10] However, for high-LET radiation, many types of DNA damage are complex (ie, clusters containing mixtures of two or more various types of damage, such as single-strand breaks and double-strand breaks), and occur within a localised region of DNA. Complex damage is uncommon for endogenous damage (eg, DNA damage caused by cellular errors during duplication) or low-LET radiation and has been associated with the increased relative biological effectiveness of densely ionising radiation. Figure 3 compares charged-particle tracks visualised in nuclear emulsions[14] with patterns of DNA double-strand-break distributions in human cells. DNA double-strand breaks are visualised in situ by γ-H2AX immunofluorescence staining[15]—a technique that exploits the rapid phosphorylation of the histone H2AX in the chromatin surrounding a DNA double-strand break.[16] The different patterns of energy deposition for various particles is shown in the different distribution of double-strand breaks in cells.

Repair of double-strand breaks occurs through direct end-joining and homologous recombination. Exposure to high-LET radiation, in which complex double-strand breaks occur with high frequency, seems to result in little repair, and thus cell death. Misrejoining of unrepairable ends of DNA with other radiation-induced double-strand breaks leads to large DNA deletions and chromosome aberrations. The high effectiveness in cell killing is the rationale for heavy-ion cancer treatment—ie, hadrontherapy;[17] radiotherapy with high-energy charged nuclei rather than x-rays—but residual damage in surviving cells is of concern for carcinogenesis.

Heavily charged particles effectively produce inter-chromosomal exchanges, with **relative biological effectiveness** exceeding 30 in interphase (as visualised by use of premature chromosome condensation) for energetic iron ions.[18] The detailed association between relative biological effectiveness and LET found for total chromosome exchanges is consistent with previous studies of relative biological effectiveness versus LET with respect to mutation[19] and to in-vitro neoplastic transformation in rat cells.[20] For all these endpoints, relative biological effectiveness peaks at about 100–200 keV/μm and decreases at very high LET. However, the nature of chromosome damage is different with heavy ions compared with that of sparsely ionising radiation. Large differences in gene expression are noted on exposure to x-rays compared with that for HZE ions, showing differences in damage-response pathways.[21,22] Furthermore, qualitative differences between x-rays and HZE in the type of gene mutations have been reported.[23] Multicolour fluorescence-painting techniques of chromosomes of human beings have shown that high-LET α-particles[24] and iron ions[25,26] induce substantially more complex chromosome exchanges in cells than does low-
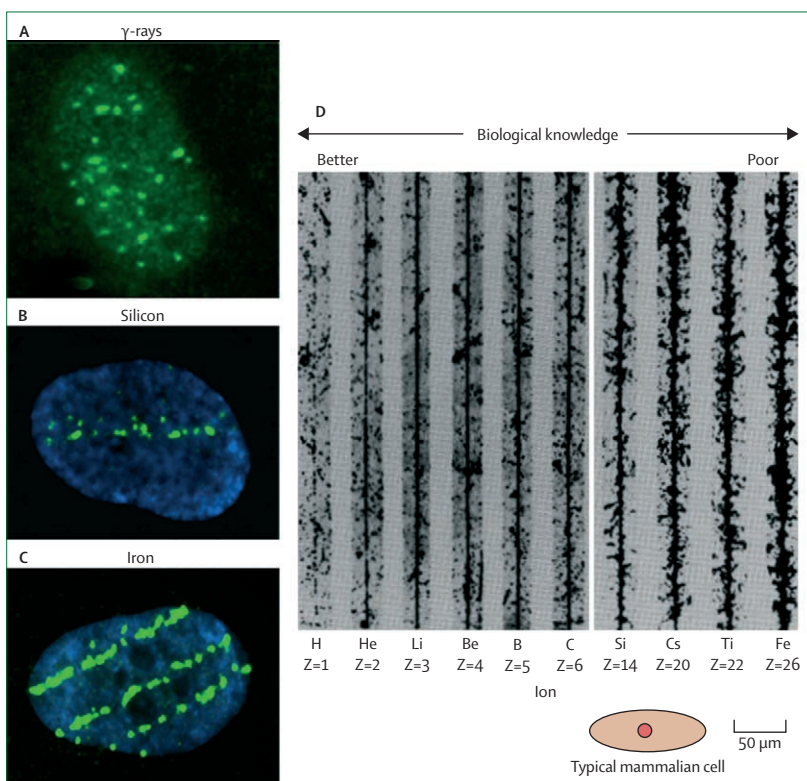

Figure 3: Comparison of particle tracks in nuclear emulsions and human cells
Three nuclei of human fibroblasts exposed to (A) γ-rays, (B) silicon ions, or (C) iron ions; and immunostained for detection of γ-H2AX.[15] Every green focus corresponds to a DNA double-strand break. In the cell exposed to sparsely ionising γ-rays (A), H2AX foci are uniformly distributed in the nucleus. Cells exposed to HZE particles show DNA damage along tracks—one silicon (B) and three iron (C) particles, respectively. Spacing between DNA double-strand breaks is reduced at very high-LET. (D) Tracks of different ions, from protons to iron, in nuclear emulsions,[14] show increasing ionisation density (LET=ΔE/Δx) as charge, Z, increases. Biological knowledge increases with increasing atomic number.

LET radiation. Most of these complex chromosome rearrangements ultimately lead to cell death. Only a small amount of the initial cytogenetic damage is measured in mice 2–4 months after exposure to energetic iron ions.[27] A low relative biological effectiveness for the induction of late chromosome damage has been measured in the progeny of human lymphocytes exposed in vitro to energetic iron ions. However, terminal deletions (eg, single truncated chromosomes) occur at much higher frequency in the progeny of cells exposed to heavy ions compared with those exposed to γ-rays.[28]

Presence of chromosomes without telomeres in the progeny of cells exposed to heavy ions is of particular interest. Sabatier and colleagues[29] found that rearrangements of telomere regions were associated with chromosome instability in human fibroblasts, many generations after exposure to accelerated heavy ions. Telomere dysfunction has a crucial part in initiating or sustaining genomic instability,[30,31] which is an important step in cancer progression. The effects induced by heavy-ions on telomere stability have been studied by use of small-interfering RNA knockdown of components of

Relative biological effectiveness
The ratio of doses of γ-rays and heavy ions that produce the same effect

DNA-dependent protein kinases in human lymphoblasts.[32] Exposure of cells with reduced DNA-dependent protein kinase expression to γ-rays or iron particles shows differential effects on telomere dysfunction, mutation frequency, and differential effects between radiation qualities. Different results were found for γ-rays and HZE nuclei: iron nuclei were more effective in producing double-strand breaks—telomere fusions when DNA-dependent protein kinases are inactive. Cells containing telomere-deficient chromosomes either senesce or undergo cycles of breakage, fusion, and bridging cycles, promoting genetic instability. The fate of normal cells containing one truncated chromosome with loss of the telomere is not known, but loss of one telomere in cancer cells can result in instability in several chromosomes.[33] These results suggest that telomere instability is an important early event in the pathway to cancer induction by HZE nuclei.

Radiobiology of HZE ions: tissue effects
Studies in animals generally show that HZE nuclei have a higher carcinogenic effect than does low-LET radiation. Relative biological effectiveness factors comparing γ-rays with HZE ions in mice or rats for tumours of the skin[34] and of the Harderian[35] or mammary[36] glands recorded values as high as 25–40 at low doses. However, the risk of developing cancer, and its effect on factors such as quality of life, cannot be characterised fully until the relation between radiation quality and latency, in which tumours develop earlier after high-LET irradiation than after that of low LET,[37] is defined adequately. The short latency and increased effectiveness noted for HZE ions compared with sparsley ionising radiation was similar to that of previous studies of neutrons,[37] and, together with the lack of response for tumour induction in mice by γ-rays recorded in many low-dose studies, suggests that the ideas for scaling used in present risk-assessment approaches are unable to define important qualitative effects and that relative biological effectiveness factors are potentially not definable or are faulty.

Studies have discussed the importance of DNA damage and mutation or extracellular-matrix remodelling and other non-targeted effects as initiators of carcinogenesis.[38] Tissue effects independent of DNA damage associated with cancer initiation or progression include genomic instability,[39] extracellular-matrix remodelling,[38] persistent inflammation,[38] and oxidative damage.[40] Other studies are investigating: the relations between radiation, activation of dormant tumours, and modulation of angiogenesis;[41] the acceleration of non-cancer risks that occur during aging, including cataracts;[42] and damage to the CNS.[43,44]

So-called bystander or non-targeted effects[40,45] could have important consequences for space exploration. These effects occur in cells that are not hit directly by an ionising particle, but which are affected by signals from irradiated cells in tissue up to about 1 mm away.[46] These long-range effects in tissues are important in assessment of the risk exposure to low-dose radiation. Non-targeted effects might increase the risk at low doses, thus reducing the effectiveness of spacecraft shielding; however, they might have a protective effect if cells surrounding the hit cell are removed by apoptosis from the organism. Both potential effects challenge the conventional ==linear no-threshold-risk model== assumption,[47] which is used for radioprotection on Earth and in space, but is still disputed in the scientific community.[48] Moreover, these effects suggest important targets for biological countermeasures likely to be more effective than those targeting DNA damage.[49]

Conclusion
Radiation-induced cancer is one of the main health risks for manned exploration of the Solar system. Epidemiological studies on Earth have shown that exposure to moderate to high doses of ionising radiation increases the risk of cancer in most organs. Leukaemia and cancers of the breast, thyroid, colon, and lung are particularly sensitive to induction by radiation.[50] However, risk uncertainties for space radiation tumorigenesis are still very high because the radiation quality in space is very different from that on Earth. Reduction of the uncertainties in risk assessment, which are needed before a mission to Mars, has led to many investigations guided by molecular and genetic research on carcinogenesis and degenerative diseases. The main uncertainties in risk-projection models will be reduced only by improvement of basic understanding of the underlying biological processes and their disruption by space radiation. Unique features are involved in this approach because of the specific challenges to biological systems presented by space radiation, especially HZE ions. The issue of radiation risk during space exploration is unlikely to be solved by a simple countermeasure, such as shielding or radioprotective drugs. The risk will be understood and controlled only with further basic research in cancer induction by charged particles.

References
1 White RJ, Averner M. Humans in space. *Nature* 2001; **409:** 1115–18.
2 US National Council for Radiation Protection and Measurements. Radiation protection guidance for activities in low-Earth orbit. Report number 132. Bethesda, MD, USA: National Council for Radiation Protection and Measurements, 2000.
3 Cucinotta FA, Schimmerling W, Wilson JW, et al. Space radiation cancer risks and uncertainties for Mars missions. *Radiat Res* 2001; **156:** 682–88.
4 Wilson JW, Thibeault SA, Cucinotta FA, et al. Issues in protection from galactic cosmic rays. *Radiat Environ Biophys* 1995; **34:** 217–22.
5 Cucinotta FA, Nikjoo H, Goodhead DT. The effects of delta rays on the number of particle-track transversals per cell in laboratory and space exposures. *Radiat Res* 1998; **150:** 115–19.

6   Saganti PB, Cucinotta FA, Wilson JW, et al. Radiation climate map for analyzing risks to astronauts on the Mars surface from galactic cosmic rays. *Space Sci Rev* 2004; **110:** 143–56.

7   Preston DL, Shimizu Y, Pierce DA, et al. Studies of mortality of atomic bomb survivors. Report 13: solid cancer and noncancer disease mortality: 1950–1997. *Radiat Res* 2003; **160:** 381–407.

8   Goodhead DT. Initial events in the cellular effects of ionizing radiations: clustered damage in DNA. *Int J Radiat Biol* 1994; **65:** 7–17.

9   Todd P. Stochastics of HZE-induced microlesions. *Adv Space Res* 1989; **9:** 31–34.

10 Blakely EA, Kronenberg A. Heavy-ion radiobiology: new approaches to delineate mechanisms underlying enhanced biological effectiveness. *Radiat Res* 1998; **150** (suppl 5): S126–45.

11 Sutherland BM, Bennett PV, Sidorkina O, Laval J. Clustered DNA damages induced in isolated DNA and in human cells by low doses of ionizing radiation. *Proc Natl Acad Sci USA* 2000; **97:** 103–08.

12 Gulston M, De Lara C, Jenner T, et al. Processing of clustered DNA damage generates additional DSB in mammalian cells post-irradiation. *Nucleic Acids Res* 2004; **32:** 1602–09.

13 Rydberg B, Cooper B, Cooper PK, et al. Dose-dependent misrejoining of radiation-induced DNA double-strand breaks in human fibroblasts: experimental and theoretical study for high- and low-LET radiation. *Radiat Res* 2005; **163:** 526–34.

14 McDonald FB. Review of galactic and solar cosmic rays. In: Arthur Reetz Jr, ed. Second symposium on protection against radiations in space. NASA Special Publication-71. Washington, DC: NASA, 1965: 19–29.

15 Desai N, Davis E, O'Neill P, et al. Immunofluorescence detection of clustered gamma-H2AX foci induced by HZE-particle radiation. *Radiat Res* 2005; **164:** 518–22.

16 Celeste A, Fernandez-Capetillo O, Kruhlak MJ, et al. Histone H2AX phosphorylation is dispensable for the initial recognition of DNA breaks. *Nat Cell Biol* 2003; **5:** 675–79.

17 Amaldi U, Kraft G. Radiotherapy with beams of carbon ions. *Rep Prog Phys* 2005; **68:** 1861–82.

18 George K, Durante M, Willingham V, et al. Biological effectiveness of accelerated particles for the induction of chromosome damage measured in metaphase and interphase human lymphocytes. *Radiat Res* 2003; **160:** 425–35.

19 Kiefer J. Mutagenic effects of heavy charged particles. *J Radiat Res (Tokyo)* 2002; **43** (suppl): S21–25.

20 Yang TCH, Craise LM, Mei MT, Tobias CA. Neoplastic cell transformation by heavy charged particles. *Radiat Res Suppl* 1985; **8:** 177–87.

21 Ding L, Shingyoji M, Chen F, et al. Gene expression changes in normal human skin fibroblasts induced by HZE-particle radiation. *Radiat Res* 2005; **164:** 523–26.

22 Chang PY, Bjornstad KA, Rosen CJ, et al. Effects of iron ions, protons and X rays on human lens cell differentiation. *Radiat Res* 2005; **164:** 531–39.

23 Kronenberg A, Gauny S, Criddle K, et al. Heavy ion mutagenesis: linear energy transfer effects and genetic linkage. *Radiat Environ Biophys* 1995; **34:** 73–78.

24 Anderson RM, Stevens DL, Goodhead DT. M-FISH analysis shows that complex chromosome aberrations induced by alpha-particle tracks are cumulative products of localized rearrangements. *Proc Natl Acad Sci USA* 2002; **99:** 12167–72.

25 Durante M, George K, Wu H, Cucinotta FA. Karyotypes of human lymphocytes exposed to high-energy iron ions. *Radiat Res* 2002; **158:** 581–90.

26 Johannes C, Horstmann M, Durante M, et al. Chromosome intrachanges and interchanges detected by multicolor banding in lymphocytes: searching for clastogen signatures in the human genome. *Radiat Res* 2004; **161:** 540–48.

27 Tucker JD, Marples B, Ramsey MJ, Lutze-Mann LH. Persistence of chromosome aberrations in mice acutely exposed to $^{56}Fe^{+26}$ ions. *Radiat Res* 2004; **161:** 648–55.

28 Durante M, George K, Cucinotta FA. Chromosomes lacking telomeres are present in the progeny of human lymphocytes exposed to heavy ions. *Radiat Res* 2006; **165:** 51–58.

29 Sabatier L, Dutrillaux B, Martin MB. Chromosomal instability. *Nature* 1992; **357:** 548.

30 Maser RS, DePinho. Connecting chromosomes, crisis, and cancer. *Science* 2002; **297:** 565–69.

31 Feldser DM, Hackett JA, Greider CW. Telomere dysfunction and the initiation of genome instability. *Nat Rev Cancer* 2003; **3:** 623–27.

32 Zhang Q, Williams, ES, Askin K, et al. Suppression of DNA-PK by RNAi has different quantitative effects on telomere dysfunction and mutagenesis in human lymphoblasts treated with gamma rays or HZE particles. *Radiat Res* 2005; **164:** 497–504.

33 Sabatier L, Ricoul M, Pottier G, Murnane JP. The loss of a single telomere can result in instability of multiple chromosomes in a human tumor cell line. *Mol Cancer Res* 2005; **3:** 139–50.

34 Burns FJ, Jin Y, Koenig KL, Hosselet S. The low carcinogenicity of electron radiation relative to argon ions in rat skin. *Radiat Res* 1993; **135:** 178–88.

35 Alpen EL, Powers-Risius P, Curtis SB, DeGuzman R. Tumorigenic potential of high-Z, high-LET charged-particle radiations. *Radiat Res* 1993; **136:** 382–91.

36 Dicello JF, Christian A, Cucinotta FA, et al. In vivo mammary tumorigenesis in the Sprague-Dawley rat and microdosimetric correlates. *Phys Med Biol* 2004; **49:** 3817–30.

37 Fry RJM, Storer JB. External radiation carcinogenesis. *Adv Radiat Biol* 1987; **13:** 31–91.

38 Barcellos-Hoff MH, Park C, Wright EG. Radiation and the microenvironment—tumorigenesis and therapy. *Nat Rev Cancer* 2005; **5:** 867–75.

39 Park CC, Henshall-Powell RL, Erickson AC, et al. Ionizing radiation induces heritable disruption of epithelial cell interactions. *Proc Natl Acad Sci USA* 2003; **100:** 10728–33.

40 Mothersill C, Seymour CB. Radiation-induced bystander effects: implications for cancer. *Nat Rev Cancer* 2004; **4:** 158–64.

41 Folkman J, Watson K, Ingber D, Hanahan D. Induction of angiogenesis during the transition from hyperplasia to neoplasia. *Nature* 1989; **339:** 58–61.

42 Cucinotta FA, Manuel FK, Jones J, et al. Space radiation and cataracts in astronauts. *Radiat Res* 2001; **156:** 460–66.

43 Joseph JA, Hunt WA, Rabin BM, Dalton TK. Possible "accelerated striatal aging" induced by ^{56}Fe heavy particle irradiation: implications for manned space flights. *Radiat Res* 1992; **130:** 88–95.

44 Rola R, Sarrkissan V, Obenaus A, et al. High-LET radiation induced inflammation and persistent changes in markers of hippocampal neurogenesis. *Radiat Res* 2005; **165:** 556–60.

45 Prise KM, Schettino G, Folkard M, Held KD. New insights on cell death from radiation exposure. *Lancet Oncol* 2005; **6:** 520–28.

46 Belyakov OV, Mitchell SA, Parikh D, et al. Biological effects in unirradiated human tissue induced by radiation damage up to 1 mm away. *Proc Natl Acad Sci USA* 2005; **102:** 14203–08.

47 Brenner DJ, Doll R, Goodhead DT, et al. Cancer risks attributable to low doses of ionizing radiation: assessing what we really know. *Proc Natl Acad Sci USA* 2003; **100:** 13761–66.

48 Tubiana M Aurengo A, Averbecj D, Masse R. Recent reports on the effect of low doses of inoizing radiation and its dose-effect relationship. *Radiat Environ Biophys* 2006; **44:** 245–51.

49 Coleman C, Stone HB, Moulder JE, Pellmar TC. Modulation of radiation injury. *Science* 2004; **304:** 693–94.

50 Wakeford R. The cancer epidemiology of radiation. *Oncogene* 2004; **23:** 6404–28.